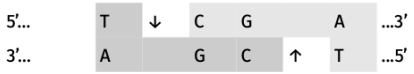
GeneSpin Srl Via Friuli, 51 -20135 Milano
P.IVA e C.F. 04520270960
administration@pec.genespin.com
info@genespin.com

DATA SHEET

info@genespin.com - www.genespin.com

TaqI

cat. no	Units
STS-TaqI	3500 Units
	5 x 3500 Units



Unit Definition: One unit is the amount of enzyme required to completely digest 1 µg of Lambda DNA (dam-; 121 sites) in 1 hour in a total reaction volume of 50 µl. Enzyme activity was determined in the recommended reaction buffer.

For general laboratory use.

Shipping: shipped on gel packs
Storage Conditions: store at -20 °C
Additional Storage Conditions: avoid freeze/thaw cycles
Shelf Life: 12 months
Form: liquid (Supplied in 10 mM Tris-HCl pH 7.4, 300 mM KCl, 0.1 mM EDTA, 1 mM DTT, 500 µg/ml BSA and 50 % [v/v] glycerol)

Concentration: 10 units/µl

Source: Thermus aquaticus YT I
Supplied with: 10x Universal Buffer (UB)

Recommended 50 µl assay

10* µl	10x Universal Buffer (UB)
1 µg	pure DNA[1] or PCR product[2]
10 units	enzyme
fill up to 50 µl	PCR grade water

Optimum buffer condition for restriction is 2x UB
[1] Supercoiled or high molecular weight DNA (e.g. plant genomic DNA) may require longer incubation time or higher amount of enzyme.
[2] Some enzymes may require additional DNA bases flanking the restriction site for complete digestion.

Protocol:
• The enzyme should not exceed 10 % of total reaction volume.
• Add enzyme as last component. Mix components well before adding enzyme. After enzyme addition, mix gently by pipetting.
Do not vortex.
• Incubate 5 to 10 min. at 60 °C.
• Stop reaction by alternatively:
- Addition of 2.1 µl EDTA pH 8.0 [0.5 M], final 20 mM
- Heat Inactivation (No)
- Spin Column DNA Purification
- Gel Electrophoresis and Single Band Excision
- Phenol-Chloroform Extraction or Ethanol Precipitation.

Double Digestion - Buffer Compatibility:
B1 - 10-25 % Relative Activity
B2 - 50-75 % Relative Activity
B3 - 75-100 % Relative Activity
B4 - 50-75 % Relative Activity
B5 - 50 % Relative Activity
1x UB - 100 % Relative Activity (recommended)
Please note that the optimum digestion condition for this enzyme is 1x UB. Within the Universal Buffer (UB) system, the most majority of our enzymes display 100% Relative Activity in 1x UB and only few either in 0.5x or 2x UB. If optimum condition for second enzyme is different than the recommended for the first enzyme, we suggest carrying out first the restriction at the higher recommended concentration of UB and dilute the reaction volume to the adequate UB concentration for further proceeding with the second restriction.

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Reaction Enzymes Buffer Guide:

Buffer 1:	10 × B1	
	100 mM	Tris-HCl(pH 7.9, 25°C)
	100 mM	MgCl2
	1000 µg/ml	BSA
Buffer 2	10 × B2	
	100 mM	Tris-HCl(pH 7.9, 25°C)
	100 mM	MgCl2
	500 mM	NaCl
	1000 µg/ml	BSA
Buffer 3	10 × B3	
	500 mM	Tris-HCl(pH 7.9, 25°C)
	100 mM	MgCl2
	1000 mM	NaCl
	1000 µg/ml	BSA
Buffer 4	10 × B4	
	100 mM	Tris-HCl(pH 7.9, 25°C)
	100 mM	MgCl2
	1500 mM	NaCl
	1000 µg/ml	BSA
Buffer 5	10 × B5	
	200 mM	Tris-HCl(pH 7.9, 25°C)
	100 mM	Mg acetate
	500 mM	K acetate
	1000 µg/ml	BSA

Reaction Buffer Compatibility:

Our restriction enzymes are fully compatible to restrictases and buffer systems from other manufacturers and can be used along in double digestions. To obtain best results, consult the corresponding manuals of all involved products.

DNA Methylation:

No Inhibition: dcm, CpG Inhibition (Blocked by overlapping): dam

Quality Control:

All preparations are assayed for contaminating endonuclease, 3'- exonuclease, 5' exonuclease/ 5' phosphatase, as well as nonspecific single- and doublestranded DNase activities.

Ligation and recutting:

After 10-fold overdigestion with TaqI, >90 % of the DNA fragments can be ligated and recut with this enzyme.

Note:

Incubation without BSA results in 50 % activity.